WEBINAR ON STATTISTICS FOR MICROBIOLOGISTS

AMR Data Collection, Curation, Analysis and Antibiogram Development

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By end of this presentation, you will be able to learn:







Importance and sources of AMR data Identification of data source and variables

AMR information cycle, Data collection, curation

Basic cleaning





- AMR data analysis using different tools Tools, Trends, patterns, antibiograms
- Antibiograms, Dashboard and Research Develop antibiogram and publish manuscript
- AI, Machine learning and Data science
- Drug discovery, Prediction and Mobile application

Why is AMR data important

"On the basis of our predictive statistical models, there were an estimated 4.95 million

(3.62–6.57) deaths associated with bacterial AMR in 2019, including 1.27 million deaths

attributable to bacterial AMR."



e an estimated 4·95 million including 1·27 million deaths

AMR information cycle



Source: https://www.open.edu/openlearncreate/mod

National and local level policy, planning and advocacy

Which data units, where, and how many

Sources of data and necessary information (variables)





Primary: surveys, interviews, routine record keeping, laboratory tests or experiments. **Secondary:** Existing data are accessed and analysed by someone other than the person(s) who collected the data.

VITEK 2 or MYLA Software

Manual Registers

Laboratory Information System

Excel Sheet/ Others

Example of a Sample data

| Fi | le Home | Insert | Ρ | age l | Layout | Formulas | Data | Review View | Help | (| Q 1 | Fell me | what | you v | vant to do | |
|----|----------------|--------|-----|----------------|----------|-----------|------------|---------------------|-------|------|-----|---------|------|-------|------------|-----|
| C2 | 7 🔹 | : × | ~ | f _x | 70 | | | | | | | | | | | |
| | А | | В | С | D | E | F | G | | н | 1 | J | K | L | м | N |
| 1 | Identification | number | Sex | Age | Specimen | _date Yea | r Sample | Organism | A | MC A | ٩ΜК | CIP | CRO | СТХ | GEN | SXT |
| 2 | _9242362412_ | | m | 70 | 1/2/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 5 | S | | | S | R |
| 3 | _9139642321_ | | f | 34 | 1/2/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 5 | R | | | S | S |
| 4 | _8021891052_ | | m | 52 | 1/2/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 5 | R | | | S | R |
| 5 | _1617418339_ | | m | 27 | 1/2/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 5 | S | | | S | S |
| 6 | _1501885197_ | | m | 48 | 1/2/2017 | 201 | 7 Wound Sv | Staphylococcus aure | eus R | S | 5 | R | | | S | R |
| 7 | _1301121252_ | | m | 60 | 1/2/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 6 | R | | | S | R |
| 8 | _6584123381_ | | f | 70 | 1/2/2017 | 201 | 7 Urine | Klebsiella sp. | | S | 5 | S | S | | S | R |
| 9 | _5245247211_ | | m | 60 | 1/2/2017 | 201 | 7 Urine | Klebsiella sp. | | S | 5 | R | S | | S | R |
| 10 | _5140148802_ | | f | 12 | 1/2/2017 | 201 | 7 Wound Sv | Klebsiella sp. | | R | 2 | R | R | R | R | R |
| 11 | _5120871702_ | | f | 63 | 1/2/2017 | 201 | 7 Urine | Klebsiella sp. | | R | 1 | R | R | | R | R |
| 12 | _4620205931_ | | m | 63 | 1/2/2017 | 201 | 7 Pus | Klebsiella sp. | R | S | 5 | R | R | R | S | R |
| 13 | _2442203663_ | | f | 47 | 1/2/2017 | 201 | 7 Pus | Klebsiella sp. | R | S | 5 | S | S | S | S | S |
| 14 | _1240941431_ | | m | 65 | 1/2/2017 | 201 | 7 Urine | Klebsiella sp. | | S | 5 | R | S | | S | R |
| 15 | _6328111119_ | | f | 70 | 1/2/2017 | 201 | 7 Blood | Escherichia coli | | S | 5 | S | S | | S | S |
| 16 | _4718848462_ | | f | 57 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | R | S | | S | R |
| 17 | _3955138154_ | | f | 35 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | R | S | | S | S |
| 18 | _3673267750_ | | m | 65 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | R | R | | S | S |
| 19 | _2526268228_ | | m | 58 | 1/2/2017 | 201 | 7 Pus | Escherichia coli | | R | 8 | S | S | S | R | R |
| 20 | _1641141162_ | | f | 27 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | R | S | | S | S |
| 21 | _1531292172_ | | f | 40 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | R | 2 | R | R | | R | S |
| 22 | _1516984671_ | | f | 70 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | S | S | | S | S |
| 23 | _0186641342_ | | f | 43 | 1/2/2017 | 201 | 7 Urine | Escherichia coli | | S | 5 | R | R | | R | S |
| 24 | _6783982251_ | | f | 40 | 1/3/2017 | 201 | 7 Pus | Staphylococcus aure | eus S | S | 5 | S | | | S | R |
| 25 | _6393247351_ | | f | 55 | 1/3/2017 | 201 | 7 Wound Sv | Staphylococcus aure | eus R | S | 5 | R | | | R | R |
| 26 | _3108178241_ | | m | 70 | 1/3/2017 | 201 | 7 Pus | Staphylococcus aure | eus | S | 5 | R | | | S | R |

Sources of data and necessary variables ...



Status, Hospital information, Date of

admission, Date of discharge,

Number of days previous

hospitalisation

Diabetes, Chronic Kidney Disease,

Chronic Liver Disease, CKD, Cancer,

Other

Urine, Stool, Wound swab/ pus,

Sputum, Blood, Endotracheal

aspirate and other

- Identified organizms
- Zone of inhibition value or
 - RIS result
 - Both

How is measured AMR data?

Study objective: To estimate the country/hospital level AMR prevalence and trends for WHO priority pathogens and other clinically important and frequently isolated pathogens as well as to enable the comparison of countries on spatiotemporal maps¹.

> No. of non-susceptible isolates AMR rate=

No. of tested isolates

The AMR rate is the extent to which a pathogen is resistant to a particular antimicrobial agent or class as is determined by the proportion of isolates that are non-susceptible (i.e., either intermediate or resistant) over a one-year period

X 100 (CI 95%)

** CIs for AST data can be constructed using the Wilson score method

To determine the association between AMR and its potential drivers, the following patient and country-level factors were considered:

- **Patient-level factors:** demographics (age and gender), diagnosis, comorbidities, antimicrobial usage, presence of device (catheter, central line or ventilator) and origin of infection (hospital or community)
- **Country-level factors:** Global Health Security index scores on AMR prevention, primary education, GDP per capita, physician and nurse density, disease prevalence and antibiotic consumption in defined daily dose (DDD) per 1 000 inhabitants (the country-level associations are presented separately at a regional or continental level)



AMR data analysis (Conceptual framework)

Patient demographic information

Risk factors, clinical history

Origins (Hospital or community)

Pathogens/ Organisms

Samples

Independent variables



Dependent variable

AMR data analysis (Descriptive stats)

- **Descriptive analysis for categorical variables**
 - Count: AST results showed that 38 of the 120 MRSA isolates were resistant to levofloxacin
 - Proportion: AST results showed that 31.7% of the 120 MRSA isolates were resistant to levofloxacin
 - Frequency: Found 120 positive isolates
- **Descriptive analysis for numeric variables**
 - Arithmetic mean
 - Geometric mean
 - Median
 - Mode

Measures of central tendency

- Percentiles
- Interquartile range •
- Variance
- Standard variance \bullet



AMR data analysis (Inferential stats)

- Allow us to make predictions about a population, based on data collected from a sample lacksquare
- Discuss in detail in the next session \bullet



AMR data curation and analysis tools

R-Programming

- R Package for AMR: https://msberends.github.io/ AMR
- Example: https://julhas.com/blog/amrdata-analysis-using-r



Research | Conferences | Presentations | DHIS2 Profile | DHIS2 trainingland | DHIS2 ePortal | JS Edutech | MPH Materials | QAAPT | Hiking | Travelling | Running | Blog

Back to blog

AMR Data Analysis Using R

Posted: 23 December 2 Author: Julhas Sujan

About AMR and R package

R package to simplify the analysis and prediction of Antimicrobial Resistance (AMR) and to work with microbial and antimicrobial data and properties, by using evidence-based methods. Copyright by: https://msberends.github.io/AMR/index.html#copyright

Outine

Stata/ SPSS

- Available commands
- Example: https://julhas.com/blog/amrdata-analysis-using-stata



search | Conferences | Presentations | DHIS2 Profile | DHIS2 Trainingland | DHIS2 ePortal | JS Edutech | MPH Materials | QAAPT | Hiking | Travelling | Runni

Back to blog AMR Data Analysis Using Stata

sted: 22 December 2021

About AMR and Stata

The IBM Stata software will help us to analyze the antimicrobial resistance data. We can use cross tabulation, pearson's chi-squared test, bar, pie, line, box, histogram, and regression analysis to determine frequency distributions.

Outine

Python

• Python commands



ig | Bi

CIRCA

A: Each section of the diagram represents the resistance observed in *E. coli* against the antibiotic. Size of each section is proportional to the proportion of *E. coli* resistant to the antibiotic over the study period. Antibiotics of the same class are shown in similar colors.

B: Line graphs show temporal trends of proportion of resistant *E. coli* in a clockwise direction from 2011 to 2015. **C**: Bar charts show the comparison of susceptibility to resistant strains in patients of different age groups. Moving from out to inward, bars represent proportion of resistant *E. coli* reported in children <5 years of age, young adults between 6 to 18 years, middle aged 19 to 45 years old, 45 to 65 years old patients, and elderly over 65 years of age, respectively.

D: Gender-wise comparison to susceptibility to resistant *E. coli* is shown in form bars. Outer circle and inner circle shows proportion of resistant *E. coli* isolated from women vs. men, respectively.

E: For co-resistance analysis, antibiotics belonging to the same class with same susceptibility profile for all isolates of *E. coli* were merged into a single variable.

F: Proportion of *E. coli* isolates resistant to one antimicrobial resistant to another antimicrobial are shown in the connections. The area covered by the connection on E is proportional to the level of co-resistance observed. Co-resistance proportions were scaled down to 1/15th of the actual overlap for visualization.

Source: https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0250226



Fig: Antimicrobial resistance in *Escherichia coli* (E. coli).

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| م 🥥 | Analysis | s results - Organis | sm = All organis | sms (n=42016 Isolates) | | | | | | | | | | | | | | | | | | | — | þ | \times |
|-----|----------|---------------------|------------------|-----------------------------|----------------------|-------------|--------|--------|-----|------|-----------------|---------------|--------|---|---|---|---|----|----|----|----|----|----|----|----------|
| Fi | le E | dit | | | | | | | | | | | | | | | | | | | | | | | |
| | Сор | y table | Copy grap | oh Save table | Save | graph | Cor | ntinue | | Show | / hidden column | S | | | | | | | | | | | | | |
| | | Organisms | Code | Antibiotic name | Site of infection | Breakpoints | Number | %R | %I | %S | %R 95%C.I. | %S 95%C.I. | Number | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 ^ |
| ► | A | ll organisms | CRB_ND100 | Carbenicillin | | (None) | 8146 | 78.3 | 0.7 | 21 | 77.4-79.2 | 20.1-21.9 | 0 | | | | | | | | | | | | |
| | A | ll organisms | MET_ND5 | Methicillin | | 10 - 13 | 1700 | 56.7 | 0.4 | 42.9 | 54.3-59.1 | 40.5-45.3 | 0 | | | | | | | | | | | | |
| | A | ll organisms | AMC_ND20 | Amoxicillin/Clavulanic acid | | 14 - 17 | 10240 | 89.6 | 0.2 | 10.2 | 89.0-90.2 | 9.7-10.8 | 0 | | | | | | | | | | | | |
| | A | ll organisms | TZP_ND100 | Piperacillin/Tazobactam | | 18 - 20 | 9393 | 48.5 | 2.1 | 49.4 | 47.5-49.5 | 48.4-50.4 | 1 | | | | | | | | | | | | |
| | A | ll organisms | CRO_ND30 | Ceftriaxone | | 20 - 22 | 12916 | 77 | 0.5 | 22.5 | 76.2-77.7 | 21.8-23.2 | 0 | | | | | | | | | | | | |
| | A | ll organisms | FEP_ND30 | Cefepime | | 19 - 24 | 749 | 56.5 | 0.5 | 43 | 52.8-60.0 | 39.4-46.6 | 0 | | | | | | | | | | | | |
| | A | ll organisms | IPM_ND10 | Imipenem | | 20 - 22 | 3117 | 42.4 | 2.2 | 55.3 | 40.7-44.2 | 53.6-57.1 | 1 | | | | | | | | | | | | |
| | A | ll organisms | MEM_ND10 | Meropenem | | 20 - 22 | 4997 | 46 | 1.1 | 53 | 44.6-47.4 | 51.6-54.3 | 0 | | | | | | | | | | | | |
| | A | ll organisms | AMK_ND30 | Amikacin | | 15 - 16 | 13947 | 55.3 | 0.6 | 44.1 | 54.5-56.1 | 43.2-44.9 | 0 | | | | | | | | | | | | |
| | A | ll organisms | GEN_ND10 | Gentamicin | | 13 - 14 | 13531 | 59 | 0.3 | 40.6 | 58.2-59.9 | 39.8-41.5 | 0 | | | | | | | | | | | | ~ |
| < | | | • | | - | • | | | | | | - | | | | | | | | | | | | | > |



https://whonet.org

AMASS

Section [2]: Isolate-based surveillance report

Blood: Escherichia coli

(No. of patients = 6)

| Gentamicin H |
|--|
| Amikacin 🛏 🔤 |
| Co-trimoxazole 🛏 🛶 |
| Ampicillin 🛏 🛏 🔤 |
| FLUOROQUINOLONES |
| Ciprofloxacin ——— |
| Levofloxacin |
| 3GC ⊢ |
| Cefpodoxime |
| Ceftriaxone |
| Cefotaxime |
| Ceftazidime 🛏 🛏 |
| Cefepime |
| CARBAPENEMS |
| Imipenem H |
| Meropenem |
| Ertapenem |
| Doripenem |
| Colistin |
| · · · · · · · · · · · · · · · · · · · |
| 0 20 40 60 80 100 *Proportion of NS isolates(%) |

| Antibiotic agent | Proportion of NS isolates (n) | 95% CI |
|------------------|-------------------------------|-----------|
| Gentamicin | 0% (0/6) | 0% - 39% |
| Amikacin | 0% (0/6) | 0% - 39% |
| Co-trimoxazole | 0% (0/6) | 0% - 39% |
| Ampicillin | 33% (2/6) | 10% - 70% |
| FLUOROQUINOLONES | 0% (0/6) | 0% - 39% |
| Ciprofloxacin | 0% (0/6) | 0% - 39% |
| Levofloxacin | NA | - |
| 3GC | 0% (0/6) | 0% - 39% |
| Cefpodoxime | NA | - |
| Ceftriaxone | NA | - |
| Cefotaxime | 0% (0/6) | 0% - 39% |
| Ceftazidime | 0% (0/6) | 0% - 39% |
| Cefepime | NA | - |
| CARBAPENEMS | 0% (0/6) | 0% - 39% |
| Imipenem | 0% (0/6) | 0% - 39% |
| Meropenem | NA | - |
| Ertapenem | NA | - |
| Doripenem | NA | - |
| Colistin | NA | - |

https://amass.website/

QAAPT



| Escherichia coli | • | Amoxicillin / Clavulanic ac 🔻 | 5 | • | Q | |
|------------------|---|-------------------------------|---|---|---|--|
| | | | | | | |

| CSV | | | | | Search: | | |
|--------------------|----|----------------|------------------------------|------------|-------------------|------|---|
| name | Å. | Percentage (%) | Number of Resistant Isolates | Total test | Combined Value | | |
| | | 43.64 | 1424 | 3263 | 43.64 (1424/3263) | | • |
| / Clavulanic acid | | 89.28 | 1933 | 2165 | 89.28 (1933/2165) | | |
| in | | 88.04 | 162 | 184 | 88.04 (162/184) | | |
| | | 56.06 | 37 | 66 | 56.06 (37/66) | | |
| 2 | | 77.62 | 2487 | 3204 | 77.62 (2487/3204) | | |
| in | | 69.74 | 1987 | 2849 | 69.74 (1987/2849) | | |
| | | 17.76 | 322 | 1813 | 17.76 (322/1813) | | |
| e | | 64.89 | 2114 | 3258 | 64.89 (2114/3258) | | |
| | | 52 53 | 1653 | 31/17 | 52 53 (1653/31/7) | | - |
| o 16 of 16 entries | | | | | Previous 1 | Next | |

Antimicrobial resistance patterns by year



2016
2017
2018
2019
2020

Other platforms

- ATLAS: Antimicrobial Testing Leadership And Surveillance
- http://www.bccdc.ca/health-professionals/data-reports/antimicrobial-resistance- \bullet utilization/antimicrobial-resistance-dashboard
- https://amrmap.net/
- https://dashboard.globalamrhub.org/ ullet
- https://amrhub.ru/ lacksquare
- https://amrcloud.net/en/
- https://amr.tghn.org
- https://www3.paho.org/data/index.php/en/mnu-topics/antimicrobial-resistance/572-amr-acercaulleten.html
- NARMS: https://www.fda.gov/animal-veterinary/antimicrobial-resistance/national-antimicrobial- \bullet resistance-monitoring-system

Antibiogram components

- Table 1: Distribution of bacterial growth in different specimen
- Table 2: Pattern of organisms isolated ullet
- Table 3: Distribution of bacteria isolated from OPD, IPD •
- Table 4: Distribution of sex and specimens
- Table 5: Antibiotic susceptibility pattern of major Gram-positive organisms ullet
- Table 6: Antibiotic susceptibility pattern of major Gram-negative organisms ullet
- Table 7: Distribution of organisms and specimens

Example of cumulative antibiograms

| | Less restricted antibiotics | | | | | | | | | | | | | | | | | | | | |
|---|---|-----|------|-----|----|-----|----|-----|----|-----|----|-----|----|-----|-----|-----|----|-----|----|----------|-----|
| | Less restricted antibiotics | | | | | | | | | | | | | | | | | | | | |
| | Mo. Mo. Mo. Mo. Mo. M | | | | | | | | | | | | | | | | | | | | |
| | Organism | z | %S | n | %S | n | %S | n | %S | n | %S | n | %S | n | %S | n | %S | n | %S | <u>n</u> | %S |
| | Escherichia coli | 821 | - 54 | 821 | 84 | 821 | 87 | 816 | 77 | 821 | 79 | 801 | 96 | 821 | 97 | 821 | 76 | 802 | 92 | 53 | 100 |
| | Klebsiella preum oniae | 133 | R | | 89 | 133 | 85 | 131 | 84 | 133 | 88 | 130 | | | 95 | 133 | 88 | 130 | 98 | 118 | 99 |
| Pseudom onas aeruginosa 105 R R R R 92 105 60 104 95 97 95 Enterococcus faecalis 80 100 80 R R R 100 80 I | | | | | | | | | | | | | | 95 | | | | | | | |
| Enterococcus faecalis 80 100 80 R 100 80 I <th< td=""><td></td></th<> | | | | | | | | | | | | | | | | | | | | | |
| Proteus mirabilis 58 93 58 100 58 86 58 90 58 91 58 R 98 58 100 58 100 54 10 ESCAPPM group 68 R 79 68 81 67 93 68 75 68 79 62 10 | | | | | | | | | | | | | | | 100 | | | | | | |
| ESCAPPM group 68 R 79 68 81 67 93 68 75 68 79 62 10 | | | | | | | | | | | | | | 100 | | | | | | | |
| <70% of is plates sensitive 70-89% of is plates sensitive >90% of is plates sensitive not tested or not clinically effective antibiption R intrinsic resistance <i>Ecoli</i> ESBL = 2.8% of is plates <i>K pneumonia</i> ESBL = 10.5% of is plates CRE-1 is plate of <i>Bnterobacter cloacae Enterococc uss p</i> -85 is plates <i>States (VRE = 0% of is plates) Staphylococc us pureus</i> - 27 is plates (MRSA = 26% of is plates) | | | | | | | | | | | | | | | | | | | | | |
| NOTE 1. Data processed by OrgTRx (antibiogram)s oftware to exclude multiple is plates so only the first is plate of a given species per patient per year per 2. Only organisms with greater than 30 is plates are included (CLSI Guideline M39-A2 recommends that results should include at least 30 is plates to 3. ESCAPPM group includes <i>Enterobacter, Serratia, Otrobacter</i> (excluding <i>C. koseri</i>), <i>Aeromonas, Hafnia, Providencia, Pontoea</i> and <i>Morganella</i> species per 4. Antimicrobial susceptibility testing method: EUCAST microbroth dilution and disc diffusion | | | | | | | | | | | | | | | | | | | | | |

Source: https://www.safetyandquality.gov.au/sites/default/files/migrated/A-Specification-for-Hospital-Cumulative-Antibiograms-December-2013.pdf

| | | R | estri | cted | Antik | piotic | s | | |
|----------|-----|-----------------|-----------|----------|----------------|-------------|-----|--------------|-----------|
| Amiharin | | المطابقة محمدهم | centexote | Mercenet | iliai ado aixi | Northersein | | a janaaaaa M | vancompan |
| s | п | %s | n | ŝ | n | %s | n | %s | п |
| 00 | 803 | 96 | 804 | 100 | 804 | 94 | 306 | R | |
| 99 | 130 | 88 | 130 | 100 | 130 | 85 | 130 | R | |
| 95 | 100 | R | | 91 | 105 | 90 | 100 | R | |
| | | R | | | | | | 100 | 79 |
| 00 | 58 | 100 | 58 | 100 | 58 | 100 | 58 | R | |
| 00 | 68 | 81 | 68 | 99 | 68 | 90 | 68 | R | |

ic not recommended to be used in children without specialist advice

er subtype (eg. urine) is included to be considered significant. pp.

Example of cumulative antibiograms ...

| Hospital Antibiogram – 2019 Inpatient – All Units Period: 01/01/19 – 12/31/19 | | TOTAL ISOLATES | Penicillin | Ampicillin | Ampicillin/sulbactam | Piperacillin/tazobactam | Ceftriaxone | Cefepime | Meropenem | Levofloxacin | Oxacillin | SXT | Nitrofurantoin | Gentamicin | Vancomycin |
|---|--------------------------|----------------|-------------------------|------------|----------------------|-------------------------|-------------------------|----------|-----------|--------------|-----------|-----|----------------|------------|------------|
| ÷ | Escherichia coli | 2215 | | 41 | 54 | 94 | 85 | 88 | 100 | 77 | | 76 | 97 | 91 | |
| Gram (–) | Klebsiella pneumoniae | 532 | | | 72 | 95 | 89 | 94 | 99 | 91 | | 83 | 35 | 93 | |
| | Pseudomonas aeruginosa | 446 | | | | 92 | | 91 | 87 | 81 | | | | 90 | |
| | Staphylococcus aureus | 821 | | | | | | | | | 62 | 95 | | | 100 |
| 0 | MRSA | 312 | | | | | | | | | 0 | 94 | | | 100 |
| am (+ | MSSA | 509 | | | | | | | | | 100 | 96 | | | 100 |
| Ū | Streptococcus pneumoniae | 47 | 97 (83) ^a | | | | 97 (86) ^a | | | 100 | | | | | 100 |
| | Enterococcus spp. | 295 | | 87 | | | | | | | | | 99 | | 89 |

Antibiogram by R

| $\langle \neg \neg \rangle$ | 🖅 🔚 🖸 Source on Save 🔍 🎢 🖌 📃 |
|-----------------------------|--|
| 1 | # AMR works great with dplyr, but it's not required or neccesary |
| 2 | #Julhas Aug 02, 2023 |
| 3 | library(AMR) |
| 4 | library(dplyr) |
| 5 | |
| 6 | example_isolates %>% |
| 7 | <pre>mutate(bacteria = mo_fullname()) %>%</pre> |
| 8 | <pre># filtering functions for microorganisms:</pre> |
| 9 | filter(mo_is_gram_negative(), |
| 10 | <pre>mo_is_intrinsic_resistant(ab = "cefotax")) %>%</pre> |
| 11 | # antibiotic selectors: |
| 12 | select(bacteria, |
| 13 | aminoglycosides(), |
| 14 | carbapenems()) |
| 15 | antibiogram(example_isolates, |
| 11:26 | (Top Level) \$ |
| | |
| Console | Terminal × Background Jobs × |

😱 R 4.2.2 · ~/ 🖗

i For aminoglycosides() using columns 'GEN' (gentamicin), 'TOB' (tobramycin), 'AMK' (amikacin), and 'KAN' (kanamycin) i For carbapenems() using columns 'IPM' (imipenem) and 'MEM' (meropenem) # \triangle tibble: 35 x 7

| # A LIDDIE. 33 X / | | | | | | | |
|--|---|---------------|-------------|-------------|-------------|-------------|-----|
| bacteria | GEN | TOB | AMK | KAN | IPM | MEM | |
| <chr></chr> | <sir< td=""><td>> <sir></sir></td><td><sir></sir></td><td><sir></sir></td><td><sir></sir></td><td><sir></sir></td><td></td></sir<> | > <sir></sir> | <sir></sir> | <sir></sir> | <sir></sir> | <sir></sir> | |
| 1 Pseudomonas aeruginosa | I | S | NA | R | S | NA | |
| 2 Pseudomonas aeruginosa | I | S | NA | R | S | NA | |
| 3 Pseudomonas aeruginosa | I | S | NA | R | S | NA | |
| 4 Pseudomonas aeruginosa | S | S | S | R | NA | S | |
| 5 Pseudomonas aeruginosa | S | S | S | R | S | S | |
| 6 Pseudomonas aeruginosa | S | S | S | R | S | S | |
| 7 Stenotrophomonas maltophi | lia R | R | R | R | R | R | |
| 8 Pseudomonas aeruginosa | S | S | S | R | NA | S | |
| 9 Pseudomonas aeruginosa | S | S | S | R | NA | S | |
| 10 Pseudomonas aeruginosa | S | S | S | R | S | S | |
| # i 25 more rows | | | | | | | |
| <pre># i Use `print(n =)` to s</pre> | see more | rows | | | | | |
| > antibiogram(example_isolat | es, | | | | | | |
| + antibiotics = | c(aminog | lycosid | es(), (| carbap | enems () |))) | |
| i Using column 'mo' as input | for col. | _mo. | | | | | |
| i For aminoglycosides() using | g columns | GEN' | (genta | umicin) | , 'тое | ' (tobran | iyc |
| i For carbapenems() using co | lumns 'I | PM' (imi | ipenem) | and ' | MEM' (| meropenen | i) |
| i 502 combinations had less t | than min | imum = 3 | 0 resu | lts an | d were | ignored | |
| # A tibble: 10×7 | | | | | | | |
| `Pathogen (N min-max)` | AMK GE | N IPM | KAN | MEM | TOB | | |
| * <chr> <a< td=""><td>lb1> <db1< td=""><td>> <db1></db1></td><td><db1></db1></td><td><db1></db1></td><td><db1></db1></td><td></td><td></td></db1<></td></a<></chr> | lb1> <db1< td=""><td>> <db1></db1></td><td><db1></db1></td><td><db1></db1></td><td><db1></db1></td><td></td><td></td></db1<> | > <db1></db1> | <db1></db1> | <db1></db1> | <db1></db1> | | |
| 1 CONS (43-309) | 0 8 | 6 52 | 0 | 52 | 22 | | |
| 2 E. coli (0-462) | 100 9 | 8 100 | NA | 100 | 97 | | |
| 3 E. faecalis (0-39) | 0 | 0 100 | 0 | NA | 0 | | |
| 4 κ. pneumoniae (0-58) | NA 9 | 0 100 | NA | 100 | 90 | | |
| 5 P. aeruginosa (17-30) | NA 10 | 0 NA | 0 | NA | 100 | | |
| 6 P. mirabilis (0-34) | NA 9 | 4 94 | NA | NA | 94 | | |
| 7 S. aureus (2-233) | NA 9 | 9 NA | NA | NA | 98 | | |
| 8 5. epidermidis (8-163) | 0 7 | 9 NA | 0 | NA | 51 | | |
| 9 5. hominis (3-80) | NA 9 | 2 NA | NA | NA | 85 | | |

cin), 'AMK' (amikacin), and 'KAN' (kanamycin)

| | - Pa | atnogen (N min-max) | AMK | GEN | TBW | KAN | MEM | TOB | |
|----|------|---------------------|-------------|-------------|-------------|-------------|-------------|-------------|--|
| * | <0 | hr> | <db1></db1> | <db1></db1> | <db1></db1> | <db1></db1> | <db1></db1> | <db1></db1> | |
| 1 | Col | NS (43-309) | 0 | 86 | 52 | 0 | 52 | 22 | |
| 2 | Ε. | coli (0-462) | 100 | 98 | 100 | NA | 100 | 97 | |
| 3 | Ε. | faecalis (0-39) | 0 | 0 | 100 | 0 | NA | 0 | |
| 4 | к. | pneumoniae (0-58) | NA | 90 | 100 | NA | 100 | 90 | |
| 5 | Ρ. | aeruginosa (17-30) | NA | 100 | NA | 0 | NA | 100 | |
| 6 | Ρ. | mirabilis (0-34) | NA | 94 | 94 | NA | NA | 94 | |
| 7 | s. | aureus (2-233) | NA | 99 | NA | NA | NA | 98 | |
| 8 | s. | epidermidis (8-163) | 0 | 79 | NA | 0 | NA | 51 | |
| 9 | s. | hominis (3-80) | NA | 92 | NA | NA | NA | 85 | |
| 10 | s. | pneumoniae (11-117) | 0 | 0 | NA | 0 | NA | 0 | |

>





Artificial Intelligence, Machine Learning and Data Science

2010

2020

Year



0%·



(n = 1001, model: binomial)



Next: Descriptive and inferential statistics

| Parametric or non- parametric? | Outcome variable | Number of groups <mark>1</mark> | Statistical test | K |
|-----------------------------------|--|------------------------------------|---|----|
| Parametric | Categorical: nominal with two levels (dichotomous) | Two or more | Chi-squared test | E |
| Non-parametric | Categorical: ordinal, or numeric when assumptions for a t-test are not met | Two groups | Mann-Whitney U test (Wilcoxon rank-sum test) | |
| Non-parametric | Categorical: ordinal, or numeric when ANOVA test assumptions are not met | Three or more groups | Kruskal-Wallis test | 0 |
| Parametric | Numeric | Two groups | Student's t-test | |
| | | | | |
| Parametric | Numeric | Two or more groups | One-way ANOVA | |
| Parametric | Numeric | Two or more groups | Simple linear regression with one exposure variable | |
| | | | | |
| | | | | |
| Parametric | Categorical: nominal with two levels (dichotomous) | Two groups | Binomial logistic regression | Li |

Source: https://www.open.edu/openlearncreate/mod/oucontent/view.php?id=174276&printable=1

Key assumptions

Expected frequency in any cell of a contingency table is not <5 or no nore than 80% of cells have a value of <5

- Row and column totals are fixed
- Outcome can be ranked

Dutcome can be ranked

- Normal distribution of outcome variable
- Residuals have normal distribution
- Variance is the same in both groups (otherwise use modified ttest)
- Normal distribution of outcome variable
- Variance is the same in all groups
- Normal distribution of outcome variable for a given exposure value
- Linear relationship (roughly) between exposure and outcome (check with scatterplot)
- Homoscedasticity: the variance of residuals is the same for any value of the exposure variable

inear relationship between the exposure and log odds

THANK YOU

If you have any questions and queries, I will be happy to answer them during the QA session.

Feel free drop an email with your query to: info@jaetech.co

